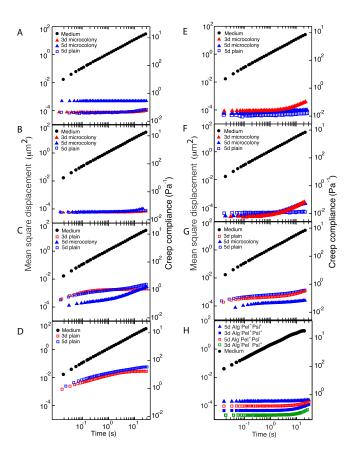


Erratum for "Dynamic Remodeling of Microbial Biofilms by Functionally Distinct Exopolysaccharides"

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Jolume 5, no. 4, doi:10.1128/mBio.01536-14, 2014. We here correct Fig. 5 (PDF page 6) with revised scaling for the right y axis. There was erroneous scaling of the creep compliance axis of Fig. 5, which affected the conversion of MSD to creep compliance values. The biological outcomes and the conclusions remain unchanged.



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FIG 5 The left panels (A to D) show MSDs (left axis) and creep compliances (right axis) of 1.0-μm particles in biofilms expressing the Alg+ Pel+ Psl+ strain (A), which is elastic and for which microcolonies are reduced in effective cross-linking from days 3 to 5, the Alg+ Pel- Psl+ strain (B), which is elastic and does not change in rheology from days 3 to 5, and the Alg+ Pel+ Psl- strain (C), which is viscoelastic and mainly consists of plains that do not change in rheology from days 3 to 5. (D) Biofilm is not formed in Alg+ Pel- Psl- cells, and particle diffusion appears to be confined by extracellular secretion from a thin layer of cells. The right panels (E to G) show MSDs and creep compliances of 0.5-μm particles in biofilms expressing the strains shown. (E) The Alg+ Pel+ Psl+ strain is elastic, and the diffusivity of particles increases at long time scales in 3-day microcolonies, indicating that the biofilm mesh size exceeds 0.5 μm. By day 5, mesh size reduces and rheology is similar to that in Alg+ Pel+ Psl- microcolonies. (F) The Alg+ Pel- Psl+ strain is elastic, and the diffusivity of particles increases at long time scales in 3- and 5-day microcolonies. The rheological properties of the microcolonies remain constant from days 3 to 5. (G) The Alg+ Pel+ Psl- strain is viscoelastic and mainly consists of plains that do not change in rheology from days 3 to 5. The 0.5-μm particles are not retained by the Alg+ Pel- Psl- cell layer. The lower curves of 0.5-μm compared to 1.0-μm particles indicate that the smaller particles locate to regions of higher effective cross-linking. (H) MSDs of 1.0-μm particles in Alg- Pel- Psl+, Alg- Pel- Psl+, and Alg- Pel- Psl- microcolonies.

We also correct the creep compliances in the text:

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A value of 4.3 \times 10^{-2} \, \text{Pa}^{-1} and not 4.3 \times 10^{-4} \, \text{Pa}^{-1} (PDF page 4, column 2, line 6). A value of 2.5 \times 10^{-1} \, \text{Pa}^{-1} and not 2.8 \times 10^{-3} \, \text{Pa}^{-1} (PDF page 4, column 2, line 10). A value of 3.6 \times 10^{-2} \, \text{Pa}^{-1} and not 3.6 \times 10^{-4} \, \text{Pa}^{-1} (PDF page 5, column 1, line 6). A value of 3.2 \times 10^{-2} \, \text{Pa}^{-1} and not 3.2 \times 10^{-4} \, \text{Pa}^{-1} (PDF page 5, column 1, line 7). A value of 1.0 \, \text{Pa}^{-1} and not 1.1 \times 10^{-2} \, \text{Pa}^{-1} (PDF page 5, column 1, line 19).
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In the Materials and Methods section (PDF page 9), in the equation for MSD conversion to creep compliance, parameter a is particle radius, not diameter.

We also correct Fig. 6 (PDF page 7). Fig. 6D was mislabeled 6A, and Fig. 6A was mislabeled 6D.

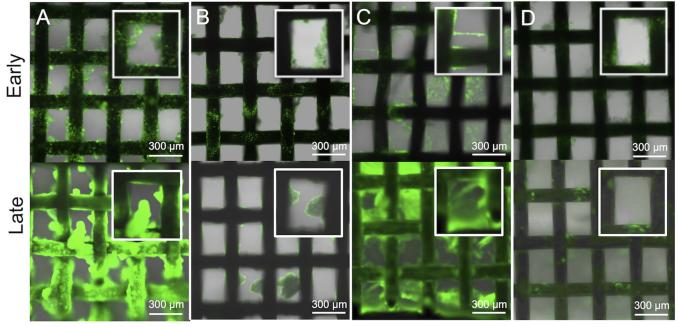


FIG 6 Confocal images of the various EPS mutants in the biofilm streamer cultivation system. Flow direction is horizontal (left to right) and parallel to the steel mesh. (A) Alg⁺ Pel⁺ Psl⁺ cells initially forming rough surface-attached biofilms that become smooth with enhanced spreading. (B) Alg⁺ Pel⁻ Psl⁺ cells forming rough surface-attached biofilms that develop large microcolonies with minimal spreading. (C) Alg⁺ Pel⁺ Psl⁻ cells forming smooth biofilms with extensive streamer formation that extends across the mesh. (D) Alg⁺ Pel⁻ Psl⁻ cells do not form biofilm. Insets show enlarged views of the biofilms.